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LA Hot Start Master Mix

(Catalogue number L608, L609, L610, L610xl)

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Description

LA Hot Start Master Mix is optimized mix for universal use in PCR. It contains a blend of two DNA polymerases, deoxyribonucleotide triphosphates (dNTPs), Dye, anti-Taq monoclonal antibody, buffer, stabilizers, additives and other components. This product is the most advanced variant of Master mix for PCR:

Rapid samples preparation, 500 µl aliquots

• 2x concentrated Master Mix is in 500 μl aliquots. This decreases possibility of contamination and allows rapid preparation of the samples by adding just specific oligonucleotide primers, template DNA and H₂O (included).

LA (Long and Accurate)

• The Mix contains unique blend of two thermostable DNA polymerases. One of them is Taq DNA polymerase, which is highly processive but lacks a 3'->5' exonuclease proofreading activity, which accounts for relatively high error rate during DNA amplification. Second polymerase is less processive but possesses 3'->5' exonuclease activity necessary for proofreading. The polymerases mix decreases error rate during synthesis of complementary DNA strands and preserves high speed of DNA synthesis. The repair capability allows amplification of complex genomic DNA fragments (up to 15 kbps in size) and less complex viral DNA (up to 40 kbps in size) and significantly decreases error rate when compared to Taq DNA polymerase

Hot Start

The Mix contains monoclonal antibody, which binds DNA polymerases and thereby blocks their enzymatic
activity and amplification of nonspecific DNA fragments. After the first denaturation cycle, the antibody is
irreversibly inactivated and Taq DNA polymerase regains enzymatic activity.

Direct loading into the gel

- The Mix contains additives and a dye, which allow direct loading of the samples into the gel without necessity of adding loading buffer.
- Dye present in the Mix migrates in the agarose gel in front of the primers and therefore does not interfere
 with quantification of the PCR products. The dye and other additives have no effect on DNA amplification
 during PCR.

High efficiency and specificity

• The Mixt allows highly sensitive and specific amplification of corresponding fragments of DNA or cDNA; it possesses MgCl₂ at a concentration suitable for most PCRs.

Technical data

Components and packaging

- 1 tube with 0.5 ml LA Hot Start Master Mix (for 40 reactions, 25 μl each).
- 1 tube with 1.5 ml PCR H₂O.

Composition

2x concentrated LA Hot Start Master Mix contains: 150 mM Tris-HCl, pH 8.8 (at 25°C), 40 mM (NH₄)₂SO₄, 0.02% Tween 20, 5 mM MgCl₂, 400 μM dATP, 400 μM dCTP, 400 μM dGTP, 400 μM dTTP, 100 U/ml Taq DNA polymerase blend, monoclonal antibody anti-Tag (38 nM), dye, stabilizers and additives.

Storage

• At temperature -20°C ± 5°C; shortly (up to one week) at temperature 4 - 30°C. This decreases demands for transportation (Nature friendly). Material can be repeatedly defrosted.

Purity and quality control

- Purity of Taq DNA polymerases is verified electrophoretically (SDS PAGE). Material is free of nucleases.
- Each batch of LA Hot Start Master Mix is tested for amplification of a single copy gene in genomic DNA.

Cat. No	Product name and specification	Quantity
L608	LA Hot Start Master Mix (1x)	40 reactions
L609	LA Hot Start Master Mix (5x)	200 reactions
L610	LA Hot Start Master Mix (25x)	1000 reactions
L610xl	LA Hot Start Master Mix (100x)	4x 1000 reactions



Protocol

Suggested basic protocol for PCR amplification using LA Hot Start Master Mix

1. In a thin-walled PCR tube the following components are mixed:

Volume*	Reagent	Final concentration
12.5 μΙ	LA Hot Start Master Mix	1x LA Hot Start Master Mix (75 mM Tris-HCl, pH 8.8, 20 mM (NH ₄) ₂ SO ₄ , 0.01% Tween 20, 2.5 mM MgCl ₂ , 200 μM dATP, 200 μM dCTP, 200 μM dTTP, 50 U/ml blend of DNA polymerases, 19 nM anti-Taq monoclonal antibody)
1 μΙ	5' primer	0.1 - 1 μM (~ 20 bases)
1 μl	3' primer	0.1 - 1 μM (~ 20 bases)
1 μl	Template DNA	
9.5 μΙ	PCR H ₂ O	to a final volume 25 μl

^{*}Different volumes can be used, but Master Mix should be finally diluted twice.

- 2. Mix gently and briefly centrifuge.
- 3. Add 20 μ l of PCR oil (Cat. No. P043) to prevent evaporation (this is not required if thermal cycler with a heated lid is used).
- 4. Perform PCR under conditions optimized for the primers used. Common cycling parameters are:

	Temperature	Time	Number of cycles
Initial denaturation	94°C	1 min	1
Denaturation	94°C	15 s	
Primers annealing	55-68°C ¹	15 s	25-35
Extension	72°C	1 min per 1 kb	
Final extension	72°C	7 min	1
Cooling	22°C		

¹Should be determined experimentally; usually 5°C below melting temperature of the primers.

5. Amplified DNA can be directly loaded into agarose gel without adding loading buffer.